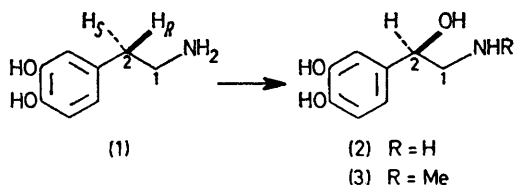


## Stereochemical Course of Hydroxylation of Dopamine by Dopamine- $\beta$ -hydroxylase (EC1.14.17.1)

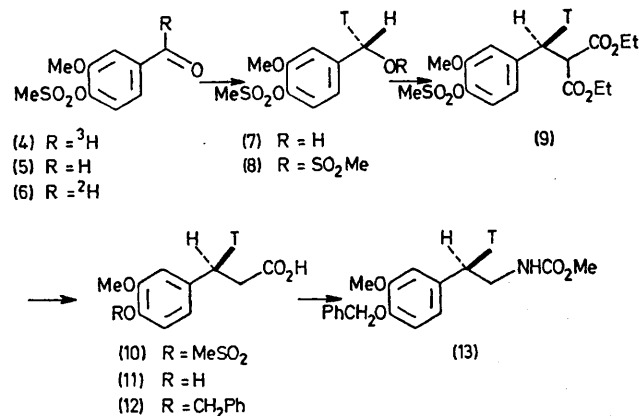
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**Summary** (2*R*)-, and (2*S*)-[2-<sup>3</sup>H<sub>1</sub>]Dopamines are synthesised having high configurational purity; they are used to prove that hydroxylation of dopamine to yield noradrenalin occurs with retention of configuration.

DOPAMINE  $\beta$ -HYDROXYLASE is a mixed function oxidase<sup>1</sup> which converts dopamine (1) into (-)-noradrenaline (2); the latter is the biological precursor of (-)-adrenaline (3) and all three catecholamines are important substances in the chemistry of the central nervous system.<sup>2</sup> Earlier work has established the illustrated (*R*)-configurations<sup>3</sup> for (-)-(2) and (-)-(3) and has shown that (a) the oxygen atom introduced at C-2 of dopamine is derived from molecular oxygen not water<sup>4</sup> and (b) only one hydrogen atom is removed from C-2 in the oxidative step.<sup>5</sup> The following studies, based on syntheses of (2*R*)-, and (2*S*)-[2-<sup>3</sup>H<sub>1</sub>] dopamines, define the stereochemistry of the hydroxylation process.



*O*-Methanesulphonyl[formyl-<sup>3</sup>H]vanillin (4) was reduced with liver alcohol dehydrogenase, its coenzyme, and ethanol to yield the (*S*)-alcohol (7) (Scheme). The corresponding dimethanesulphonyl derivative (8) was then treated with diethyl malonate anion in the presence of excess diethyl malonate to yield the diester (9). Clean S<sub>N</sub>2 inversion (see



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as the acid (14) under conditions identical with those used above. Its enantiomer (15) was prepared as before by transfer of deuterium in the initial enzymic step. Degradation of the acids (14) and (15) by ozone and then peracid yielded (2*S*)-, and (2*R*)-[2-<sup>3</sup>H<sub>1</sub>]succinic acid, respectively; both were shown to be enantiomerically pure, within experimental error, by mass spectrometric and o.r.d.<sup>8</sup> measurements. The assigned absolute configurations for the tritium labelled dopamines and their high configurational purity were thus confirmed.

TABLE. Enzymic hydroxylation of labelled dopamines

Configuration of [2- <sup>3</sup> H <sub>1</sub> ]dopamine (as 1)	<sup>3</sup> H: <sup>14</sup> C Ratio in dopamine	Conversion <sup>a</sup> (%)	<sup>3</sup> H: <sup>14</sup> C Ratio in (-)-noradrenaline <sup>b</sup>	Retention of <sup>3</sup> H (%)
(2 <i>R</i> )	14.2 ± 0.6	0.8	0.07 ± 0.02	0.5
(2 <i>RS</i> )	16.9 ± 0.6	0.6	8.3 ± 0.2	49 ± 2
(2 <i>S</i> )	13.9 ± 0.3	0.6	13.5 ± 0.3	97 ± 3

\* Conversions of ca. 1% are normal in the homogenate system.

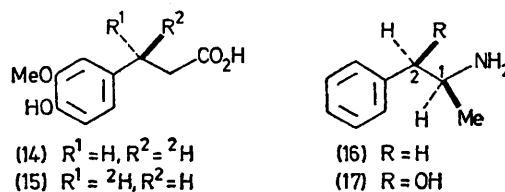
<sup>b</sup> Assayed as its tetraisobutyryl derivative; multiple determinations on repeatedly purified samples.

later) was achieved only when the strongly electron withdrawing aryl-*O*-methanesulphonyl group was used. Acid hydrolysis of the ester (9) afforded the acid (10) which *via* the phenol (11) gave the ether (12). Curtius degradation<sup>6</sup> to the urethane (13) and removal of all three protecting groups<sup>†</sup> first using alkali, and then HBr, gave (2*R*)-[2-<sup>3</sup>H<sub>1</sub>]dopamine [(1), <sup>3</sup>H at H<sub>R</sub>] isolated as its hydrochloride.

The enantiomeric product was prepared in a complementary way by reduction of unlabelled aldehyde (5) with alcohol dehydrogenase, NAD<sup>+</sup> and [1-<sup>3</sup>H]cyclohexanol.<sup>7</sup> The product [enantiomer of (7)] was then converted into (2*S*)-[2-<sup>3</sup>H<sub>1</sub>]dopamine [(1), <sup>3</sup>H at H<sub>S</sub>] by the above sequence.

<sup>†</sup> The *O*-methyl group was used because labelled 3'-*O*-methyl dopamines were required for another biosynthetic study.

The two chirally tritiated dopamines and also randomly labelled dopamine (*i.e.* 2*RS*) were mixed in known ratio with [1-<sup>14</sup>C]dopamine; the three samples were treated separately with an homogenate from bovine adrenal glands



containing ascorbic acid and proniazid (as inhibitor of amine oxidase).<sup>9</sup> Radio-inactive dopamine (**1**) and (-)-noradrenalin (**2**) were then added as carriers and isolated as their tri-isobutyryl derivatives. The results in the Table show that the hydroxylation catalysed by dopamine  $\beta$ -hydroxylase involves stereospecific removal of the 2-*pro-R* hydrogen atom from dopamine (**1**). This corresponds to retention of configuration in an attack at a benzylic carbon.

Dopamine- $\beta$ -hydroxylase will also transform several phenethylamines which are not its natural substrates; an example is (1*S*)-amphetamine (**16**) which yields (1*S*, 2*R*)-

ephedrine (**17**). Taylor has recently shown<sup>10</sup> using (1*S*, 2*S*)-[2-<sup>3</sup>H<sub>1</sub>]amphetamine (as **16**) that the hydroxylation occurs with retention of configuration which matches our finding on dopamine and also the results for biological hydroxylations in other systems.<sup>11</sup>

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